# Progesterone as a neuroactive neurosteroid, with special reference to the effect of progesterone on myelination

# E.E.Baulieu<sup>1,2,3</sup> and M.Schumacher<sup>1</sup>

<sup>1</sup>INSERM U 488, 80, rue du Général Leclerc, 94276 Le Kremlin-Bicêtre, France and <sup>2</sup>Collège de France.

<sup>3</sup>To whom correspondence should be addressed

Some steroids are synthesized within the central and peripheral nervous system, mostly by glial cells. These are known as neurosteroids. In the brain, certain neurosteroids have been shown to act directly on the function of membrane receptors for neurotransmitters. For example, progesterone inhibits the neuronal nicotinic acetylcholine receptor, whereas its 3α,5αreduced metabolite 3α,5α-tetrahydroprogesterone (allopregnanolone) activates the y-aminobutyric acid receptor complex A (GABA-R<sub>A</sub>). Besides these effects, neurosteroids also regulate important glial functions such as the synthesis of myelin proteins. Thus, in cultures of glial cells prepared from neonatal rat brain, progesterone increases the number of oligodendrocytes expressing the myelin basic protein (MBP) and the 2',3'-cyclic nucleotide-3'-phosphodiesterase (CNPase). An important role for neurosteroids in myelin repair has been demonstrated in the rodent sciatic nerve, where progesterone and its direct precursor pregnenolone are synthesized by Schwann cells. After cryolesion of the male mouse sciatic nerve, blocking the local synthesis or action of progesterone impairs remyelination of the regenerating axons, whereas administration of progesterone to the lesion site promotes the formation of new myelin sheaths.

Key words: myelination/neurosteroid/progesterone/Schwann cells/sciatic nerve

### Sex steroids: effects on the nervous system

The nervous system is a target for sex steroids (McEwen *et al.*, 1995; Priest and Pfaff, 1995). During sensitive periods in late fetal and early postnatal life, gonadal steroids influence the survival, the differentiation, and the connectivity of specific neuronal populations in both the brain and the spinal cord (De Vries *et al.*, 1984). At this stage of development, the nervous tissue is highly plastic and some of the hormone effects are permanent. In the adult, sex steroids still influence neuronal functions, mainly by regulating synaptic transmission. They do so either by increasing the transcription of specific genes after binding to intracellular receptors

(McEwen, 1991; Truss et al., 1995), or by acting directly on the neuronal membrane, most probably by binding to membrane receptors for neurotransmitters (Chadwick and Widdows, 1990; Schumacher, 1990; Majewska, 1992; Baulieu and Robel, 1995). Because of the widespread distribution of hormone-sensitive neurotransmitter receptors, such as the type A γ-aminobutyric acid (GABA<sub>A</sub>) receptor, sex steroids influence neuronal activity within large parts of the nervous system, where they exert a variety of effects that are not necessarily related to reproduction. However, this does not imply that all the effects sex steroids exert outside the brain regions involved in reproduction, such as the hypothalamus, are mediated by membrane sites. In fact, with the advent of sensitive biochemical and immunocytochemical techniques, it has become obvious that intracellular receptors for sex steroids are more widely distributed throughout the nervous system than has been thought. Thus, oestrogen receptors have been detected recently in nerve growth factor- (NGF)-sensitive neurons of the basal forebrain and dorsal root ganglia (DRG) (Toran-Allerand et al., 1992; Sohrabji et al., 1994). In addition, work in our laboratory has shown that receptors of sex steroids, such as oestradiol and progesterone, are not only present in neurons, but also in glial cells, where they mediate steroid effects on important glial functions (Jung-Testas et al., 1991, 1994).

Research performed over the past few years has shown that sex steroids continue to exert neurotrophic effects in the adult nervous system, by influencing changes in the morphology and connections of nerve cells. Indeed, despite the apparent appearance of stability, we now know that there is a continuous remodelling of neuronal connections within adult nervous tissues. For example, androgens increase the number of synaptic contacts in a group of spinal motor neurons (Matsumoto et al., 1988), and oestrogens increase the density of dendritic spines of hypothalamic and hippocampal neurons (McEwen et al., 1995). These observations have very recently stimulated research to exploit the trophic effects of sex steroids as therapeutic agents in neuronal injury. In these studies, testosterone has been identified as a trophic factor for facial and sciatic motor neurons. Indeed, this 'male sex steroid' accelerates motor neuron axonal regeneration following crush injury of the hamster hypoglossal and the rat sciatic nerve (Jones, 1993). The 'female sex steroid' progesterone has been shown to prevent death of facial motor neurons after nerve transection (Yu, 1989). These observations suggest that sex steroids may be therapeutically useful in activating or accelerating the reparative response of neurons to injury.

# Neurosteroids: concept and biosynthesis

In the light of the various effects sex steroids on both the developing and adult nervous system, it was a significant finding that some steroids, called *neurosteroids*, are synthesized within the brain and peripheral nerves by glial cells (Baulieu, 1991; Robel and Baulieu, 1994). The term neurosteroid (Baulieu, 1981) does not designate a particular class of steroids, but only refers to their site of synthesis—the nervous system. Progesterone, for example, which is a

**Figure 1.** Biosynthesis of neurosteroids in glial cells. Enzymes are shown in italics. P450scc = cytochrome P450scc (scc = side-chain cleavage),  $3\beta$ -HSD =  $\Delta$ 5-3β-hydroxysteroid dehydrogenase isomerase. Aminoglutethimide and trilostane are inhibitors respectively of the P450scc and the 3β-HSD, the minus signs indicating an inhibition.

hormone produced and secreted by the ovaries and adrenal glands, is considered to be a neurosteroid if it is synthesized within the brain or peripheral nerves.

Steroidogenesis begins with the conversion of cholesterol to pregnenolone by the cytochrome P450scc (Figure 1). This enzymatic step, which takes place in the inner mitochondrial membrane, is characteristic of the steroidogenic cells of endocrine glands, including the testes, ovaries, and adrenal glands. The first indication that pregnenolone may also be synthesized within the nervous system came from the unexpected observation that its levels are much higher in the brain than in blood (Corpéchot et al., 1983). This finding could not be explained by the cerebral retention of circulating hormone, as pregnenolone persisted in the brain days after castration and adrenalectomy, and this in spite of a very rapid cerebral clearance (Robel and Baulieu, 1985). It was then shown by immunocytochemistry that the cytochrome P450scc is expressed in the white matter throughout the brain (Le Goascogne et al., 1987). The biosynthesis of pregnenolone was demonstrated by incubating newborn rat glial cells in the presence of [3H] mevalonate, a precursor of cholesterol which easily enters cells and mitochondria (Jung-Testas et al., 1989). More recently, the P450<sub>scc</sub> mRNA has been detected by reverse transcriptase–polymerase chain reaction (RT-PCR) both in the rat brain and in cultured glial cells (Mellon and Deschepper, 1993). In agreement with the predominant localization of P450<sub>scc</sub> immunoreactivity in white matter, the enzyme is present in cultured oligodendrocytes, which are the myelinating glial cells of the central nervous system (CNS) (Jung-Testas et al., 1989). However, P450<sub>scc</sub> immunoreactivity and mRNA have also been detected

in type I astrocytes (Mellon, 1994). So far, the synthesis of pregnenolone has not been demonstrated in neurons.

Pregnenolone can be converted by the  $\Delta 5$ -3 $\beta$ -hydroxysteroid dehydrogenase isomerase (3 $\beta$ -HSD) to progesterone, which in turn is metabolized successively to 5 $\alpha$ -dihydroprogesterone (5 $\alpha$ -pregnane-3,20-dione) by the 5 $\alpha$ -reductase and to 3 $\alpha$ ,5 $\alpha$ -tetrahydroprogesterone (synonyms: 3 $\alpha$ ,5 $\alpha$ -THPR, 3 $\alpha$ -hydroxy-5 $\alpha$ -pregnane-20-one, or allopregnanolone) by the 3 $\alpha$ -hydroxysteroid oxidoreductase (Figure 1). These three enzymatic reactions are present in mixed cultures of oligodendrocytes and astrocytes (Jung-Testas *et al.*, 1989) and in type I astrocytes prepared from embryonic or neonatal rat brain (Akwa *et al.*, 1993a; Kabbadj *et al.*, 1993). Whether or not oligodendrocytes have the capability to synthesize progesterone and its reduced metabolites remains to be established.

By using testosterone as a substrate, it has been shown (Celotti *et al.*, 1992) that myelin-forming oligodendrocytes possess a high  $5\alpha$ -reductase activity and that this enzyme may be incorporated into central and peripheral myelin. *In vivo*, progesterone may be the physiological substrate for this enzyme.

Among potential metabolites of pregnenolone and progesterone are the corresponding  $20\alpha/\beta$  reduced derivatives,  $7\alpha$ -hydroxypregnenolone (Akwa *et al.*, 1992), and DHEA (dehydroepiandrosterone). The mechanism of formation of the latter is still poorly understood since the key enzyme  $17\alpha$ -hydroxylase (P  $450_{17\alpha}$ ) has not been biochemically or immunologically described, even if its mRNA has been reported at an early time of development. Unusual oxydative mechanisms may be involved (Cascio *et al.*, 1998). It is puzzling that DHEA and its sulphate [possibly formed by brain sulphotransferease activity (Rajkowski *et al.*, 1997)] has been so difficult when the demonstration of its presence and persistence in the brain after adrenalectomy and gonadectomy led to the first demonstration of neurosteroids (review in Baulieu, 1997). Available results do not support that testosterone is a neurosteroid, as is also the case for corticosterone [even though the activity of a 21-hydroxylase and the mRNA of the P450<sub>11 $\beta$ </sub>, two enzymes involved in corticosterone biosynthesis, have been detected in the brain (Mellon and Deschepper, 1993)].

The physiological stimuli that regulate neurosteroid formation are still unknown. Whether second messengers or trophic factors, which stimulate steroid biosynthesis in classic steroidogenic organs (Oonk *et al.*, 1989; Magoffin *et al.*, 1990), also increase neurosteroid formation by glial cells remains to be explored. A recent study suggests a role for autocrine factors in the regulation of neurosteroid synthesis. Thus, astrocytes preferentially convert [³H] pregnenolone to [³H] progesterone at low cell density and to 7α-hydoxylated metabolites in dense cultures (Akwa *et al.*, 1993a). In addition, the synthesis of neurosteroids may be influenced by the endocrine milieu. So far, however, only the peripheral-type benzodiazepine receptor (PBR) has been shown to play a significant role in the neurosteroidogenic activation of glial cells. This receptor is found primarily on the outer mitochondrial membrane. By increasing the intramitochondrial cholesterol transport, PBR ligands stimulate pregnenolone formation in the C6-2B glioma cell line (Costa *et al.*, 1991; Papadopoulos, 1993).

# Neurosteroids: biological significance

As reported in the previous section, it is likely that pregnenolone, progesterone, and their reduced metabolites are the only steroids which can be formed *de novo* from cholesterol within the brain. An important question is the biological significance of these neurosteroids. Probably the best guess is that they may be essential for important nervous functions and that their local synthesis may protect the brain from drops in circulating steroid levels, e.g. as they occur during the oestrus cycle or during ageing. Also, blood levels of pregnenolone and progesterone are very low in males and their nervous system may rely on a local production of these steroids. In fact, as we shall see, progesterone synthesized by glial cells plays an important role during nerve regeneration in male rodents.

Over the past few years, particular attention has been paid to the actions of pregnenolone, progesterone, and their reduced metabolites on membrane receptors for neurotransmitters. Thus, progesterone inhibits the neuronal nicotinic acetylcholine receptor (Valera *et al.*, 1992) and activates hypothalamic oxytocin receptors (Schumacher *et al.*, 1990), whereas its  $5\alpha$ , $3\alpha$ -reduced metabolite,  $3\alpha$ , $5\alpha$ TH-progesterone, activates the chloride channel of the GABA<sub>A</sub> receptor complex (Majewska *et al.*, 1986). Other neurotransmitter receptors have been identified as targets for neurosteroids and these results have been extensively reviewed (Chadwick and Widdows, 1990; Robel and Baulieu, 1994).

However, although it has been demonstrated that neurosteroids such as pregnenolone or progesterone influence the activity of neurotransmitter receptors, it has not been shown that their production by oligodendrocytes or astrocytes plays an important role in regulating receptor functions through paracrine actions. The same is true for the effects of neurosteroids on neuronal functions and behaviour. In rats, the local infusion of sulphated pregnenolone, an inhibitor of the GABA<sub>A</sub> receptor, into the nucleus basalis magnocellularis enhances memory performance. Infusion of  $3\alpha,5\alpha$ ,-tetrahydroprogesterone, an activator of the GABA<sub>A</sub> receptor, has the opposite effect (Mayo et al., 1993). Pregnenolone and steroids metabolically derived from it also have memory enhancing effects in mice (Flood et al., 1992). However, as these steroids are also produced by the gonads and adrenal glands, it is not clear whether their synthesis by glial cells is significant. This could be shown by selectively blocking or increasing the synthesis of neurosteroids within specific brain regions. Recently, pregnenolone sulphate was found to be involved in the memory mechanism taking place in the hippocampus, its decrease being correlated to the decrease of memory associated with ageing (Vallée et al., 1997). Whether the level of pregnenolone is dependent on its transformation to progesterone is not yet known.

So far, little attention has been paid to the possible trophic effects of neurosteroids. As described above, steroids exert a variety of neurotrophic effects, particularly during early development, but also in the adult. Especially after the lesion of nervous tissues, locally produced neurosteroids may stimulate, by paracrine or autocrine actions, the reparative responses of neurons and glial cells. Such an important role for neurosteroids is suggested by cell culture experiments.

When added to the culture medium, neurosteroids enhance neuronal survival (Bologa *et al.*, 1987) and increase the synthesis of myelin-specific proteins by oligodendrocytes, namely, the MBP and the CNPase (Jung-Testas *et al.*, 1994). Recently, we have begun to explore the role of neurosteroids in peripheral nerve repair (see below). Our results demonstrate for the first time that the synthesis of neurosteroids by glial cells plays an important role during the regeneration of the nervous system (Koenig *et al.*, 1995).

# The rodent sciatic nerve: a system to study the trophic actions of neurosteroids

One advantage in studying neurosteroid functions in peripheral nerves is their relatively simple structure. Sensory and motor nerve fibres are associated with a single type of glial cells, the Schwann cells, which myelinate the large axons. In addition, it is easy to prepare pure cultures of Schwann cells from neonatal rat sciatic nerves or from embryonic rat dorsal root ganglia (DRG) (Brockes et al., 1979; Kleitman et al., 1991). From the latter, it is also possible to establish cultures of sensory neurons (Kleitman et al., 1991). However, the most important feature of peripheral nerves is their remarkable regenerative capacity. Following injury, axons and their myelin sheaths distal to the lesion degenerate by a process known as Wallerian degeneration (Fawcett and Keynes, 1990), leaving behind the dividing Schwann cells, which produce a large number of neurotrophic peptides. Schwann cells indeed play a crucial role during the regeneration of nerve fibres, which begins a few hours after lesion of the nerve by axotomy, local crush, or freezing. In addition, their multiplication is critical for allowing axons to regenerate across gaps. Later, Schwann cells remyelinate the regenerating axons and, under favourable conditions, the appropriate neuromuscular connections may be restored (Fawcett and Keynes, 1990; Raivich and Kreutzberg, 1993). There are thus several steps at the level of which the actions of 'sex steroids', and in particular of neurosteroids, could promote the repair of peripheral nerves: Schwann cell proliferation, the production of neurotrophic factors, axonal growth, and remyelination.

Schwann cells in culture, prepared from neonatal rat sciatic nerves, proliferate only in the presence of elevated levels of cyclic AMP (cAMP) and specific peptide growth factors (Eccleston, 1992). We have identified insulin-like growth factor I (IGF-I) as a potent mitogen for Schwann cells in the presence of serum and dibutyryl cAMP (dbcAMP) or forskolin, a reversible activator of the adenylate cyclase. Schwann cells indeed express the receptor for IGF-I, whose number increases if they are cultured for several days in the presence of forskolin (Schumacher *et al.*, 1993). By binding to IGF-I receptors, insulin also stimulates Schwann cell proliferation at micromolar concentrations. Thus, in the presence of forskolin and a high concentration of insulin, it is possible to expand rat Schwann cell cultures and to rapidly obtain pure cells in sufficient number to study the metabolism and actions of steroids.

# Schwann cells synthesize neurosteroids

Schwann cells are not only a target for circulating steroids such as oestradiol (Eccleston, 1992). Like the glial cells of the CNS, they can synthesize neurosteroids, including pregnenolone and progesterone, and their reduced metabolites. The synthesis of pregnenolone within peripheral nerves was first suggested by the high levels of this steroid found in the human sciatic nerve (Morfin et al., 1992). In sciatic nerves of adult male rats, the concentration of pregnenolone is about ten times higher than in plasma, and is not reduced 5 days after castration and adrenalectomy (Akwa et al., 1993b). In contrast to pregnenolone, the concentration of corticosterone, a steroid of adrenal origin, is much higher in plasma than in nerves, and its concentrations drop almost to zero after removal of the steroidogenic glands. As steroids are rapidly cleared from nervous tissues (Raisinghani et al., 1968; Robel and Baulieu, 1985), these observations strongly suggest a local synthesis of pregnenolone independent of glandular sources. In fact, Schwann cells prepared from neonatal rat sciatic nerves convert [3H]-25-OH cholesterol to pregnenolone (Akwa et al., 1993b), but only when cultured in the presence of micromolar concentrations of forskolin and insulin, which are both mitogen for these cells. Thus, pregnenolone may play a role during Schwann cell proliferation. Interestingly, cAMP and IGF-I, whose effects can be mimicked by insulin, also both stimulate the expression of the cytochrome P450<sub>scc</sub> in classical steroidogenic cells (Oonk *et al.*, 1989; Magoffin et al., 1990). However, even in the presence of insulin and forskolin, the rate of pregnenolone formation by Schwann cells is very low in vitro, not exceeding 25 fmol/µg DNA/24h. This suggests that other factors may be required for an optimal activity of the P450scc enzyme in glial cells, and preliminary work indicates a crucial role of molecules of axonal origins.

In sciatic nerves of adult male rats and mice, the concentration of progesterone is also 10 times higher (~10 ng/g) than in plasma, and remains high after removal of the steroidogenic endocrine glands (Koenig et al., 1995) (Figure 2). However, pure cultures of Schwann cells prepared from neonatal rat sciatic nerves do not convert (<sup>3</sup>H) pregnenolone to progesterone, even when cultured at different cell densities in the absence or presence of the mitogens insulin and forskolin (Magoffin et al., 1990; Y. Akwa, unpublished). In contrast, Schwann cells isolated from embryonic (E18) rat DRG explants, which have been co-cultured for 4 weeks with sensory neurons, produce significant amounts of (3H) progesterone and its reduced metabolites  $5\alpha$ -dihydroprogesterone and  $3\alpha$ ,  $5\alpha$ -tetraprogesterone when incubated with (<sup>3</sup>H) pregnenolone (Figure 3). We also confirmed by immunocytochemistry that Schwann cells that have been cultured for several weeks in the presence of DRG neurons contain the 3β-HSD enzyme, which converts pregnenolone to progesterone. Taken together, these results strongly suggest that neurons induce the biosynthetic pathway of progesterone in Schwann cells.

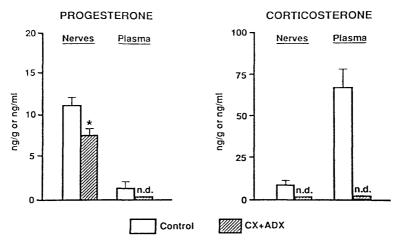


Figure 2. Levels of progesterone are ~10-fold higher in sciatic nerves than in plasma of sham-operated adult male mice (control) and remain elevated 5 days after castration and adrenalectomy (CX + ADX) (n.d. = not detectable) (mean  $\pm$  S.E.M., n = 4, \* $P \le 0.05$  when compared to controls by Student's *t*-test). In contrast, levels of corticosterone are much higher in plasma than in sciatic nerves and its levels became undetectable after CX + ADX (modified from Koenig *et al.*, 1995).

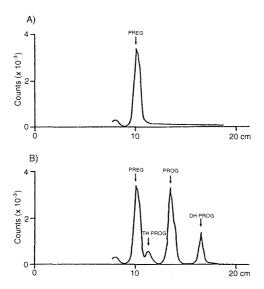


Figure 3. Metabolism of pregnenolone by rat Schwann cells in culture. The thin layer chromatograms show that (A) [ $^3$ H] pregnenolone (100 nM) is not metabolized by pure cultures of rat Schwann cells prepared from neonatal rat sciatic nerves, but (B) is converted by Schwann cells isolated from DRG explants and grown in the presence of sensory neurons for 4 weeks to [ $^3$ H] progesterone [14.3  $\pm$  0.6 (mean  $\pm$  S.E.M.) pmol/µg DNA/24 h], [ $^3$ H] 5 $\alpha$ -DH progesterone (3.9  $\pm$  0.2 pmol/µg DNA/24 h) and [ $^3$ H] 3 $\alpha$ .5 $\alpha$ -TH progesterone (2.6  $\pm$  0.1 pmol/µg DNA/24 h). [Koenig *et al.*, 1995 and A.N.Do-Thi, M.Schumacher, H.Koenig, and E.E.Baulieu, unpublished work (A)]. Preg = pregnenolone, Prog = progesterone.

Progesterone may regulate Schwann cell functions through autocrine actions. Indeed, these cells not only synthesize progesterone, they also express the intracellular receptor for this neurosteroid (progesterone-R), as shown by binding of the selective ligand (<sup>3</sup>H)ORG 2058 and by RT-PCR using primers complement-

ary to the ligand binding domain of the rat progesterone-R (I. Jung-Testas, R. Fiddes, K. Shazand, M. Schumacher, and E.E. Baulieu, unpublished work).

# Progesterone synthesized by Schwann cells promotes myelination

The production of neurosteroids is thus a feature of myelinating glial cells: oligodendrocytes in the CNS and Schwann cells in the peripheral nervous system (PNS). Therefore, we postulated that neurosteroids, and in particular progesterone, may play an important role during the process of myelination. To test this hypothesis, we examined the relationship between neurosteroids and myelin formation in the regenerating sciatic nerve of male mice after cryolesion (Koenig et al., 1995). As for other types of lesion, axons and their myelin sheaths degenerate quickly after local freezing within the distal segment of the nerve. However, the advantage of this type of lesion is that the endoneurial tubes and the Schwann cell basal lamina remain intact and guide the regenerating nerve fibres directly back to their targets. In this system, Schwann cells begin to remyelinate the regenerating axons after 1 week and the new myelin sheaths reach approximately one third of their final size after 2 weeks.

First, we showed that levels of pregnenolone and progesterone remain elevated within the regenerating mouse nerve 1 or 2 weeks after cryolesion, that is during the period of active remyelination (pregnenolone  $\sim$ 6 ng/g and progesterone  $\sim$ 10 ng/g of tissue weight). We then demonstrated that the high levels of endogenous progesterone, most probably synthesized by Schwann cells, are necessary for an efficient remyelination of the regenerating axons. In fact, local application of 100  $\mu$ g of trilostane, an inhibitor of the conversion of pregnenolone to progesterone, or of RU486, a potent competitive antagonist of progesterone, dramatically decreased the thickness of the regenerating myelin sheaths, when observed 2 weeks after lesion on electron microscopic cross-sections (Figure 4). Both inhibitors were injected at the site of lesion four times during the 2-weeks interval. The inhibitory effect of trilostane was not a toxic one since it could be reversed by the simultaneous administration of progesterone. On the other hand, repeated injections of a high dose of progesterone (100  $\mu$ g), or its direct precursor pregnenolone, significantly enhanced the number of lamellae per myelin sheath (Mellon, 1994).

It is likely that progesterone directly stimulates myelin formation by acting on Schwann cells rather than by stimulating axonal growth. This was shown by using co-cultures of DRG neurons and Schwann cells. After 4 weeks in culture, elongation of the DRG neurites is maximal and Schwann cells myelinate the sensory axons in the presence of serum and ascorbic acid (Eldridge *et al.*, 1987). At this stage of culture, myelination, but not neurite extension, was dramatically increased when a physiological concentration of progesterone (20 nM) was daily added to the culture medium for 2 weeks (Koenig *et al.*, 1995). Recently, several observations have confirmed the effects of progesterone on myelin synthesis (Desarnaud *et al.*, 1998; Melcangi *et al.*, 1998).

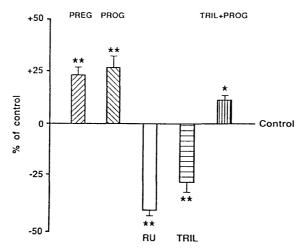


Figure 4. Role of neurosteroids in the formation of new myelin sheaths. The numbers of myelin lamellae formed 15 days after cryolesion of the right male mouse sciatic nerve were measured by electron microscopy. The number of lamellae was increased by pregnenolone and progesterone and decreased by the antiprogesterone estin RU486 (RU) and by trilostane (TRIL), an inhibitor of the conversion of pregnenolone to progesterone. The inhibitory effect of trilostane was reversed by the concomitant administration of progesterone. Steroids and their inhibitors (100  $\mu$ g in 50  $\mu$ l sesame oil) were repeatedly applied to the lesioned site immediately after surgery (day 0) and on days 5, 10 and 14. Results were expressed as percent of control (mean  $\pm$  s.e.m., n = 5, \*\* $P \le 0.01$  and \* $P \pm 0.05$  when compared to the corresponding control by Dunnett's multiple comparison tests after ANOVA) (modified from Koenig *et al.*, 1995).

# **Conclusions and perspectives**

The designation of androgens, oestrogens, and progestagens as 'sex steroids' or 'gonadal steroids' is too restrictive. First, steroids belonging to these three classes exert multiple effects on the nervous system that are not related to reproduction, including the potentiation of neuronal survival, axonal growth, and myelin formation. Second, some of these steroids are not only produced by the testes or ovaries, but are also synthesized de novo from cholesterol in the brain and peripheral nerves, mostly by glial cells. These 'neurosteroids' influence neurotransmission by acting on neurotransmitter receptors and activate important glial functions, such as myelination. Thus, progesterone synthesized by Schwann cells increases the formation of new myelin sheaths after lesion of the rodent sciatic nerve. As Schwann cells also express the intracellular receptor for progesterone, it is likely that this steroid activates the process of myelination through autocrine actions, probably by stimulating the synthesis of specific myelin proteins or lipids. From this, it follows that neurosteroids, like many peptide growth factors, belong to the important group of autocrine/paracrine factors that regulate vital functions within the nervous system. This is a significant finding, because there is much hope to use these molecules to treat diseases and injuries of the nervous system, in particular those of peripheral nerves. It is likely that neurosteroids exert their trophic effects by acting in concert, or even in synergy, with peptide growth factors such as IGF-I or nerve growth factor. The study of these interactions will become an important field of neuroendocrine research. Another critical question is the regulation of neurosteroid biosynthesis by glial cells. There are indeed two possibilities to use

neurosteroids as therapeutic agents: by stimulating their synthesis or by administering steroid compounds. Finally, an important problem that remains to be explored is the possible interactions between circulating steroids, which are secreted into the bloodstream by the gonads, and neurosteroids, which are synthesized within the nervous system. A particularly complex case is that of the role of progesterone GnRH secretion. Progesterone is not active via a classical intracellular receptor in GnRH neurons (Sterling *et al.*, 1984). Data on the direct action of progesterone, however, have been obtained recently, suggesting a membrane mechanism, possibly consistent with a postulated membrane receptor (Ramirez and Zheng, 1998). In addition, the metabolite allopregnanolone may be active through an effect at the GABA receptor (El-Etr *et al.*, 1995). This is quite a complex series of intermingled possibilities, and no doubt the complexity observed with neuroactive neurosteroids such as progesterone serve physiological purpose(s).

# Acknowledgements

This work was partially supported by the Association Française contre les Myopathies (A.F.M.) and the Mathers and Gould Foundation (New York).

#### References

- Akwa, Y., Morfin, R.F., Robel, P. and Baulieu, E.E. (1992) Neurosteroid metabolism. 7α-hydroxylation of dehydroepiandrosterone and pregnenolone by rat brain microsomes. *Biochem. J.*, **288**, 959–964.
- Akwa, Y., Sananes, N., Gouezou, M. *et al.* (1993a) Astrocytes and neurosteroids: metabolism of pregnenolone and dehydroepiandrosterone. Regulation by cell density. *J. Cell Biol.*, **121**, 135–143.
- Akwa, Y., Schumacher, M., Jung-Testas, I. and Baulieu, E.E. (1993b) Neurosteroids in rat sciatic nerves and Schwann cells. C. R. Acad. Sci. III, 316, 410–414.
- Baulieu, E.E. (1981) Steroid hormones in the brain: several mechanisms? In Fuxe, K., Gustafsson, J.A. and Wetterberg, L. (eds). *Steroid Hormone Regulation of the Brain*. Pergamon Press. Oxford, pp. 3–14.
- Baulieu, E.E. (1991) Neurosteroids: a new function in the brain. Biol. Cell, 71, 3-10.
- Baulieu, E.E. (1997) Neurosteroids: of the nervous system, by the nervous system, for the nervous system. *Recent Prog. Horm. Res.*, **52**, 1–32.
- Baulieu, E.E. and Robel, P. (1995) Non-genomic mechanisms of action of steroid hormones. In Bock, G.R. and Goode, J.A. (eds). *Non-Reproductive Actions of Sex Steroids*. Ciba Foundation Symposium 191. John Wiley & Sons, Chichester, pp. 24–42.
- Bologa, L., Sharma, J. and Roberts, E. (1987) Dehydroepiandrosterone and its sulfated derivative reduce neuronal death and enhance astrocytic differentiation in brain cell cultures. *J. Neurosci. Res.*, 17, 225–234.
- Brockes, J.P., Fields, K.L. and Raff, M.C. (1979) Studies on cultured rat Schwann cells. I. Establishment of purified populations from cultures of peripheral nerve. *Brain Res.*, **165**, 105–118.
- Cascio, C., Prasad, V.V.K., Lin, Y.Y. *et al.* (1998) Detection of P450c 17-independent pathways for dehychocpiandiesterone (DHEA) biosynthesis in brain glial tumor cells *Proc. Natl. Acad. Sci. USA*, **95**, 2862–2867.
- Celotti, F., Melcangi, R.C. and Martini, L. (1992) The 5α-reductase in the brain: molecular aspects and relation to brain function. *Front Neuroendocrinol*, **13**, 163–215
- Chadwick, D. and Widdows, K. (eds). (1990) Steroids and neuronal activity. Ciba Foundation Symposium 153. John Wiley & Sons, Chichester

- Corpéchot, C., Synguelakis, M., Talha, S. *et al.* (1983) Pregnenolone and its sulfate ester in the rat brain. *Brain Res.*, **270**, 119–125.
- Costa, E., Romeo, E., Auta, J. *et al.* (1991) Is there a pharmacology of brain steroidogenesis? In Costa, E., Paul, S.M. (eds). *Neurosteroids and Brain Function*. Thieme Medical Publishers, New York, pp. 171–176.
- De Vries, G.J., De Bruin, J.P.C., Uylings, H.B.M. and Corner, M.A. (eds). (1984) Sex differences in the brain. *Progress in Brain Research*, **61**, Elsevier, Amsterdam.
- Desarnaud, F., Do Thi An, Brown, M. et al. (1998) Progesterone stimulates the activity of the promoters of peripheral myelin protein-22 and P<sub>0</sub> genes in Schwann cells. J. Neurochem., 71, 1765.
- Eccleston, P.A. (1992) Regulation of Schwann cell proliferation: mechanisms involved in peripheral nerve development. *Exp. Cell Res.*, **199**, 1–9.
- Eldridge, C.F., Bartlett, M., Bunge, R.P. and Wood, P.M. (1987) Differentiation of axon-related Schwann cells in vitro. I. Ascorbic acid regulates basal lamina assembly and myelin formation. *J. Cell Biol.*, **105**, 1023–1034.
- El-Etr, M., Akwa, Y., Fiddes, R.J. *et al.* (1995) A progesterone metabolite stimulates the release of gonadotropin-releasing hormone from GT1-hypothalamic neurons via the γ-aminobutyric acid type A receptor. *Proc. Natl. Acad. Sci. USA*, **92**, 3769–3773.
- Fawcett, J.W. and Keynes, R.J. (1990) Peripheral nerve regeneration. Annu. Rev. Neurosci. 13, 43-60.
- Flood, J.F., Morley, J.E. and Roberts, E. (1992) Memory-enhancing effects in male mice of pregnenolone and steroids metabolically derived from it. *Proc. Natl. Acad. Sci. USA*, **89**, 1567–1571.
- Jones, K.J. (1993) Gonadal steroids and neuronal regeneration. A therapeutic role. *Adv. Neurol.*, **59**, 227–240.
- Jung-Testas, I., Hu, Z.Y., Baulieu, E.E. and Robel, P. (1989) Neurosteroids: Biosynthesis of pregnenolone and progesterone in primary cultures of rat glial cells. *Endocrinology*, 125, 2083– 2091.
- Jung-Testas, I., Renoir, J.M., Gasc, J.M. and Baulieu, E.E. (1991) Estrogen-inducible progesterone receptor in primary cultures of rat glial cells. *Exp. Cell Res.*, **193**, 12–19.
- Jung-Testas, I., Schumacher, M., Robel, P. and Baulieu, E.E. (1994) Actions of steroid hormones and growth factors on glial cells of the central and peripheral nervous system. *J. Steroid Biochem. Mol. Biol.*, **48**, 145–154.
- Kabbadj, K., El-Etr, M., Baulieu, E.E. and Robel, P. (1993) Pregnenolone metabolism in rodent embryonic neurons and astrocytes. *Glia*, **7**, 170–175.
- Kleitman, N., Wood, P.M. and Bunge, R.P. (1991) Tissue culture methods for the study of myelination. In Banker G, Goslin K (eds). *Culturing Nerve Cells*. Bradford Books, London, pp. 337–377.
- Koenig, H., Schumacher, M., Ferzaz, B. *et al.* (1995) Progesterone synthesis and myelin formation Schwann cells. *Science*, **268**, 1500–1503.
- Le Goascogne, C., Robel, P., Gouézou, M. et al. (1987) Neurosteroids: cytochrome P450scc in rat brain. *Science*, **237**, 1212–1215.
- Magoffin, D.A., Kurtz, K.M. and Erickson, G.F. (1990) Insulin-like growth factor-I selectively stimulates cholesterol side-chain cleavage expression in ovarian theca-interstitial cells. *Mol. Endocrinol.*, **4**, 489–496.
- Majewska, M.D. (1992) Neurosteroids: Endogenous bimodal modulators of the GABA<sub>A</sub> receptor. Mechanism of action and physiological significance. *Prog. Neurobiol.*, **38**, 379–395.
- Majewska, M.D., Harrison, N.L., Schwartz, R.D. *et al.* (1986) Steroid hormone metabolites are barbiturate-like modulators of the GABA receptor. *Science*, **232**, 1004–1007.
- Matsumoto, A., Micevych, P.E. and Arnold, A.P. (1988) Androgen regulates synaptic input to motor neurons of the adult rat spinal cord. *J. Neurosci.*, **8**, 4168–4176.
- Mayo, W., Dellu, F., Robel, P. et al. (1993) Infusion of neurosteroids into the nucleus basalis magnocellularis affects cognitive processes in the rat. Brain Res., 607, 324–328.
- McEwen, B.S. (1991) Steroid hormones are multifunctional messengers to the brain. *Trends Endocrinol. Metab.*, **2**, 62–67.
- McEwen, B.S., Gould, E., Orchinik, M. *et al.* (1995) Oestrogens and the structural and functional plasticity of neurons: implications for memory, ageing and neurodegenerative processes. In Bock, G.R., Goode, J.A. (eds). Ciba Foundation Symposium 191. *Non-Reproductive Actions of Sex Steroids*. John Wiley & Sons, Chichester, pp. 52–73.

- Melcangi, R.C., Magnaghi, I., Cavarretta, L. *et al.* (1998) Age-induced decrease of glycoprotein PO and myelin basic protein gene expression in the rat sciatic nerve. Repair by steroid derivatives. *Neuroscience*, **85**, 569–578.
- Mellon, S.H. (1994) Neurosteroids: Biochemistry, modes of action, and clinical relevance. *J. Clin. Endocrinol. Metab.*, **78**, 1003–1008.
- Mellon, S.H. and Deschepper, C.F. (1993) Neurosteroid biosynthesis: genes for adrenal steroidogenic enzymes are expressed in the brain. *Brain Res.*, **629**, 283–292.
- Morfin, R., Young, J., Corpéchot, C. et al. (1992) Neurosteroids: pregnenolone in human sciatic nerves. Proc. Natl. Acad. Sci. USA, 89, 6790–6793.
- Oonk, R.B., Krasnow, J.S., Beattie, W.G. and Richards, J.S. (1989) Cyclic AMP-dependent and independent regulation of cholesterol side chain cleavage cytochrome P-450 (P-450scc) in rat ovarian granulosa cells and corpora lutea. *J. Biol. Chem.*, **264**, 21934–21942.
- Papadopoulos, V. (1993) Peripheral-type benzodiazepine/diazepam binding inhibitor receptor: biological role in steroidogenic cell function. *Endocr. Rev.*, **14**, 222–240.
- Priest, C.A. and Pfaff, D.W. (1995) Actions of sex steroids on behaviours beyond reproductive reflexes. In Bock, G.R., Goode, J.A. (eds). Ciba Foundation Symposium 191. *Non-Reproductive Actions of Sex Steroids*. John Wiley & Sons, Chichester, pp. 74–89.
- Raisinghani, K.H., Dorfman, R.I., Forchielli, E. *et al.* (1968) Uptake of intravenously administered progesterone, pregnenolonenanedione and pregnenolonenanolone by the rat brain. *Acta Endocrinol.*, **57**, 393–404.
- Raivich, G. and Kreutzberg, G.W. (1993) Peripheral nerve regeneration: role of growth factors and their receptors. *Int. J. Neurosci.*, **11**, 311–324.
- Rajkowski, K.M., Robel, P. and Baulieu, E.E. (1997) Hydroxysteroid sulfotransferase activity in the rat brain and liver as a function of age and sex. *Steroids*, **62**, 427–436.
- Ramirez, V.D. and Zheng, J. (1999) Steroid receptors in brain cell membranes. In Baulieu, E.E., Robel, P. and Schumacher, M. (eds). *Neurosteroids: a New Regulatory Function in the Nervous System.* Contemporary Endocrinology Series. Totowa: Humana Press Inc. 269.
- Robel, P. and Baulieu, E.E. (1985) Neuro-steroids: 3β-hydroxy-Δ5-derivatives in the rodent brain. *Neurochem. Int.*, **7**, 953–958.
- Robel, P. and Baulieu, E.E. (1994) Neurosteroids. Biosynthesis and function. *Trends Endocrinol. Metab.*, 5, 1–8.
- Schumacher, M. (1990) Rapid membrane effects of steroid hormones: an emerging concept in neuroendocrinology. *Trends Neurosci.*, **13**, 359–362.
- Schumacher, M., Coirini, H., Pfaff, D.W. and McEwen, B.S. (1990) Behavioral effects of progesterone associated with rapid modulation of oxytocin receptors. *Science*, **250**, 691–694.
- Schumacher, M., Jung-Testas, I., Robel, P. and Baulieu, E.E. (1993) Insulin-like growth factor I: a mitogen for rat Schwann cells in the presence of elevated levels of cyclic AMP. *Glia*, **8**, 232–240.
- Sohrabji, F., Miranda, R.C. and Toran-Allerand, D.C. (1994) Estrogen differentially regulates estrogen and nerve growth factor receptor mRNAs in adult sensory neurons. *J. Neurosci.*, **14**, 459–471.
- Sterling, R.J., Gasc, J.M., Sharp, P.J. *et al.* (1984) Absence of nuclear progesterone receptor in LH releasing hormone neurones in laying hens. *J. Endocrinol.*, **102**, R5–R7.
- Toran-Allerand, C.D., Miranda, R.C., Bentham, W. et al. (1992) Estrogen receptors co-localize with low-affinity NGF receptors in cholinergic neurons of the basal forebrain. *Proc. Natl. Acad. Sci. USA*, **89**, 4668–4672.
- Truss, M., Candau, R., Chavez, S. and Beato, M. (1995) Transcriptional control by steroid hormones: the role of chromatin. In Bock, G.R., Goode, J.A. (eds). Ciba Foundation Symposium 191. *Non-Reproductive Actions of Sex Steroids*. John Wiley & Sons, Chichester, pp. 7–23.
- Valera, S., Ballivet, M. and Bertrand, D. (1992) Progesterone modulates a neuronal nicotinic acetylcholine receptor. *Proc. Natl. Acad. Sci. USA*, **89**, 9949–9953.
- Vallée, M., Mayo, W., Darnaudery, M. et al. (1997) Neurosteroids: deficient cognitive performance in aged rats depends on low pregnenolone sulfate levels in hippocampus. *Proc. Natl. Acad. Sci. USA*, **94**, 14865–14870.
- Yu, W.H. (1989) Survival of motor neurons following axotomy is enhanced by lactation or by progesterone treatment. *Brain Res.*, **491**, 379–382.